

# UNIVERSITÀ DEGLI STUDI DI MILANO-BICOCCA

# **COURSE SYLLABUS**

# **Biochemistry Practical Course**

2021-2-E0201Q052-E0201Q063M

#### **Aims**

The course aims to provide students with the theoretical and practical knowledge to manage basic experimental methodologies for biochemical analysis, through the execution of simple experiments of protein expression and analysis in mammalian cells and the discussion of results.

Knowledge and understanding. To consolidate and deepen basic knowledge of theoretical, technical and methodological issues already presented by the course of biochemistry.

Applying knowledge and understanding. To be able to correctly interpret the experimental protocols of biochemistry, recognize their salient aspects, collect and process experimental data.

Making judgments. To develop a critical vision of the experimental design and of the results achieved. To be able to recognize the context for appropriate application of the experimental procedures methods learned during the course.

Communication skills. To be able to elaborate experimental data and describe the results in an appropriate language and with the correct technical terms.

Learning skills. To be able to correctly interpret experimental protocols in contexts different from those used during the practical laboratory experience.

#### **Contents**

- 1. Manipulation and propagation of mammalian cells in vitro;
- 2. Aseptic techniques for the protection of the operator, the cell culture and the environment;

- 3. Cell growth curves and in vitro doubling time;
- 4. Use of a technique that allows the introduction of nucleic acids into mammalian cells, using a chemical method, in a process called "transfection";
- 5. Protein quantization by spectrophotometric assay;
- 6. Analysis of enzymatic activity by spectrophotometric assay;
- 7. Protein expression analysis by fluorescence microscopy;
- 8. Protein expression analysis by western blot;
- 9. Analysis and critical discussion of the results obtained from the use of the above techniques.

### **Detailed program**

- 1. In vitro manipulation and propagation of mouse and human cells, in particular will be used NIH3T3 cells either immortalized or transformed with the K.Ras oncogene;
- 2. Aseptic techniques for the protection of the operator, the product (cells) and the environment, in particular the student will be introduced to the use of a sterile biological hood, the use of sterile material and the necessary behaviors to preserve such sterility;
- 3. Cell growth curves and in vitro doubling time, in particular will be used NIH3T3 cells either immortalized or transformed with the K.Ras oncogene grown in different growth conditions;
- 4. Use of a technique that allows the introduction of nucleic acids into mammalian cells, using a chemical method, in a process called "transfection", in particular the technique of Calcium-Phosphate will be used;
- 5. Protein quantization by spectrophotometric assay, in particular the student will evaluate the protein concentration of a cell extract using the Bradford method;
- 6. Enzymatic activity analysis using a spectrophotometric assay, in particular the student will analyze the enzymatic activity of the beta-galactosidase protein;
- 7. Analysis of protein expression by fluorescence microscopy, in particular the student will analyze the expression of the Green Fluorescent Protein following its expression by transfection in mammalian cells;
- 8. Protein expression analysis by western blot technique, in particular the student will perform the complete procedure used to analyze the expression of the Green Fluorescent Protein following its expression by transfection in mammalian cells:
- 9. Critical analysis of the results obtained through the use of the aforementioned techniques.

# **Prerequisites**

Background: participation in the Biochemistry course.

Specific prerequisites: none

General prerequisites: Students can take the exams of the second year after passing the examinations of Introductory Biology, General and inorganic Chemistry, Mathematics, and Foreign Language.

# **Teaching form**

Each learning unit is addressed to a group of 35-40 students, through practical lessons which are carried out in a teaching laboratory. At the beginning of each lesson, theory, aims and experimental design will be exposed. At the end of each module, an overall discussion of collected results will take place in the same laboratory. For further details, please, refer to lesson calendar on the website of Biotechnology course.

Teaching language: italian.

# Textbook and teaching resource

Learning material (slides of introductory lessons, handout, experimental data) is available at the e-learning platform of LTA-Biochemistry module.

#### Semester

Second semester

## **Assessment method**

Two-hour written examination, consisting of six multiple choise questions and two open questions that require comprehensive and detailed answers.

#### Office hours

Contact: on demand, upon request by mail to lecturers.