



CFTR WORKFLOW

dall'arrivo del campione al referto

Corso di Laurea Magistrale in Biologia.
“Malattie genetiche: Dalla diagnosi alla terapia”

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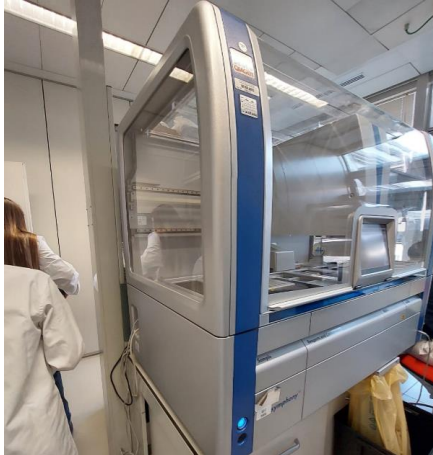
Policlinico di Milano

- Consulenza genetica
- Diagnosi prenatale
- Screening neonatale
- Diagnosi successiva al periodo neonatale



STRUTTURA LABORATORIO DI GENETICA

1. Stanza Estrazione -> accettazione del campione
2. Stanza Pre-PCR -> allestimento reazione PCR
3. Stanza Post-PCR -> reazione PCR in termociclatore



1



2



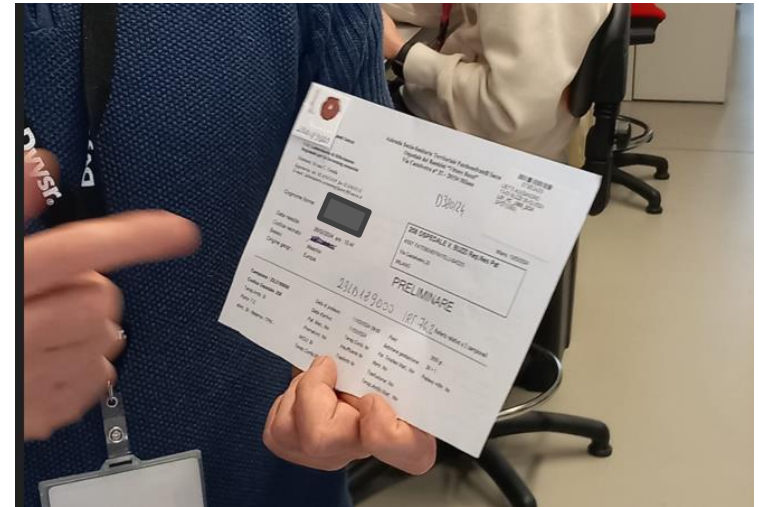
3

TIPI DI CAMPIONE

Provetta in EDTA di sangue periferico



Prelievo di sangue dal tallone di neonato

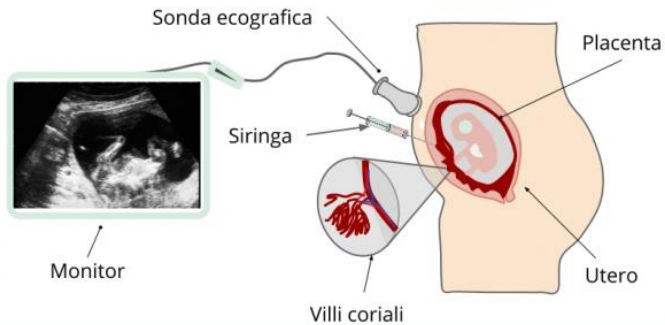


TIPI DI CAMPIONE

Villi coriali

Come si esegue la **villocentesi**?

Con un ago, sotto la guida della sonda ecografica, si raggiunge l'estremità della placenta per prelevare una piccola quantità di **villi coriali**.

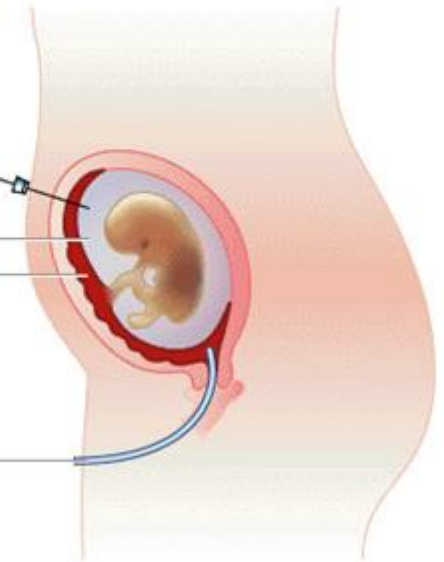


Liquido amniotico

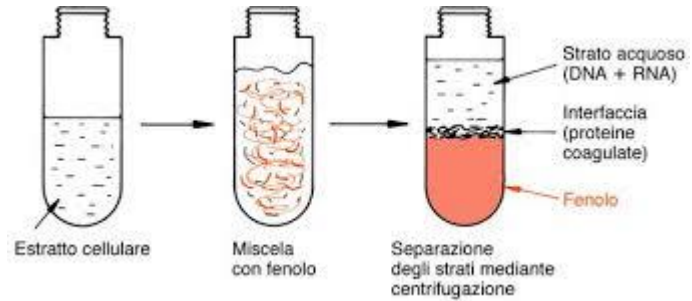
Prelievo di liquido amniotico attraverso la parete uterina (amniocentesi)

Liquido amniotico
Villi coriali

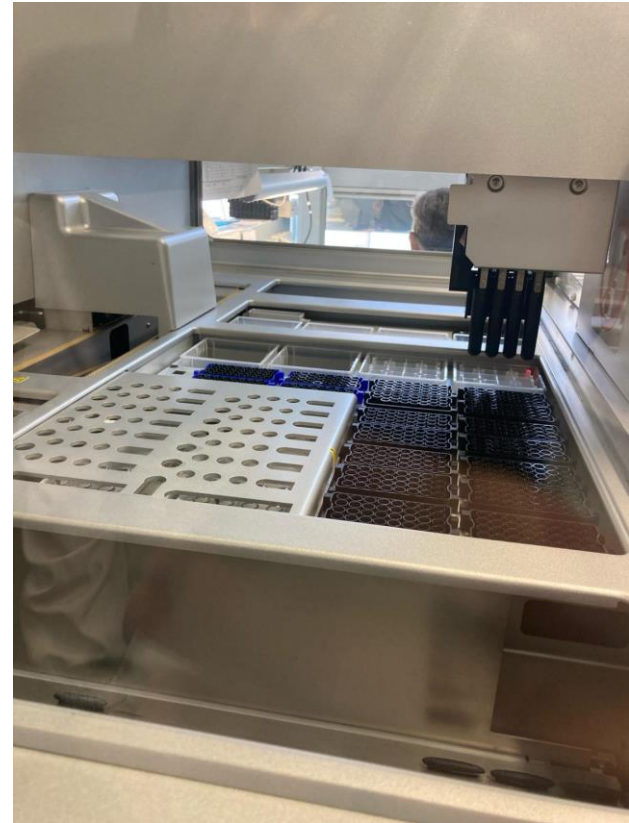
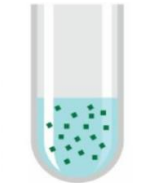
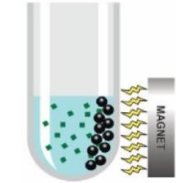
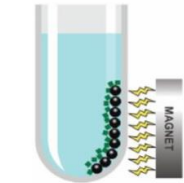
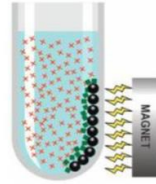
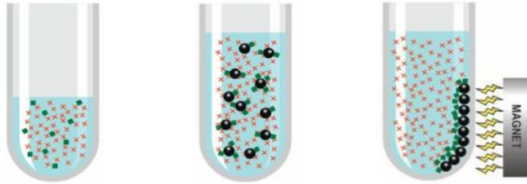
Aspirazione e prelievo dei villi coriali per via vaginale



ESTRAZIONE

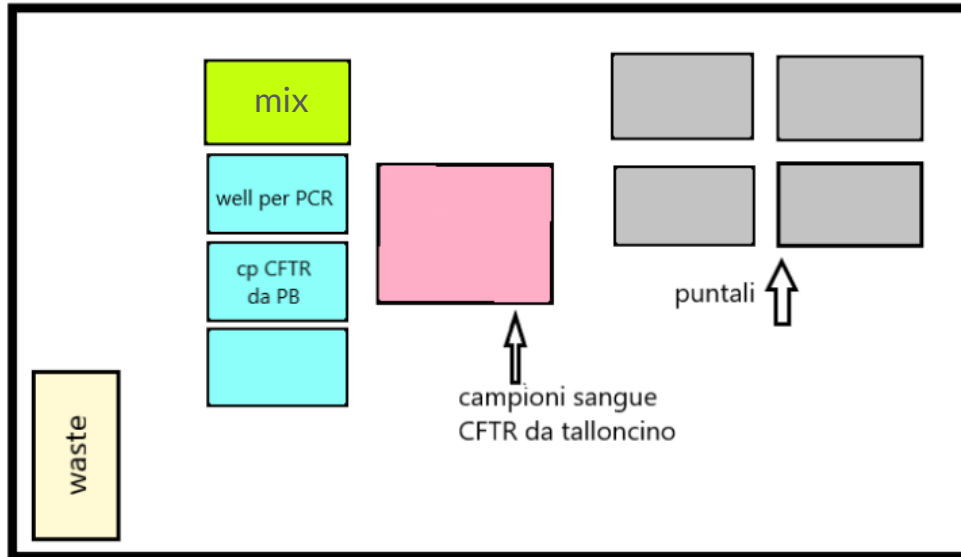


Campione di partenza Biglie legate al DNA Separazione biglie+DNA da impurità



LIQUID HANDLING

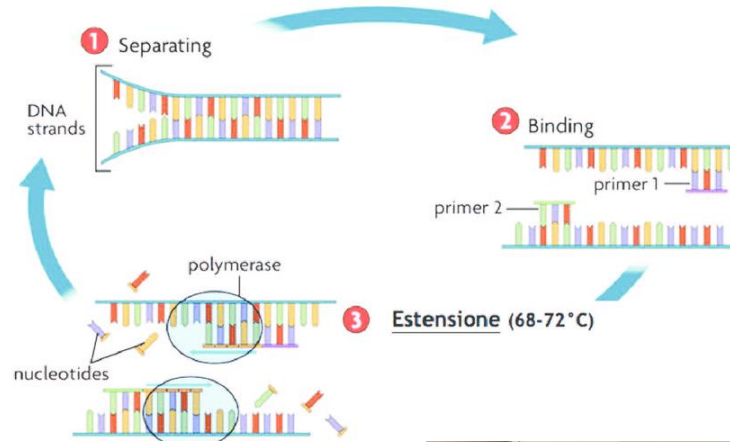
Strumento automatico per unire la mix di reazione e i campioni precedentemente diluiti.



PCR1 - amplificazione

- 22 cicli.
- 3 fasi principali:
 - Denaturazione: T = 95°C, 15 minuti.
 - Annealing: T = 62°C.
 - Estensione: T = 72°C.
- + Fase di stabilizzazione: T = 4°C.

→ Si ottengono le regioni del gene CFTR amplificate.



PCR 2 - indicizzazione

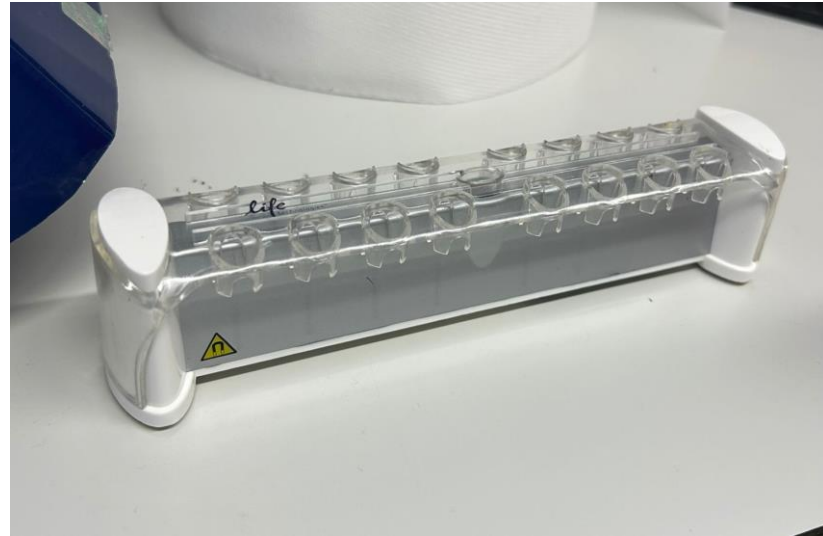
- Preparazione della mix con liquid handling.
- Utilizzo di primer che presentano all'estremità degli index specifici.
- Appaiamento dei primers alle estremità degli esoni di CFTR.

→ Si ottengono ampliconi di pazienti taggati in modo univoco.



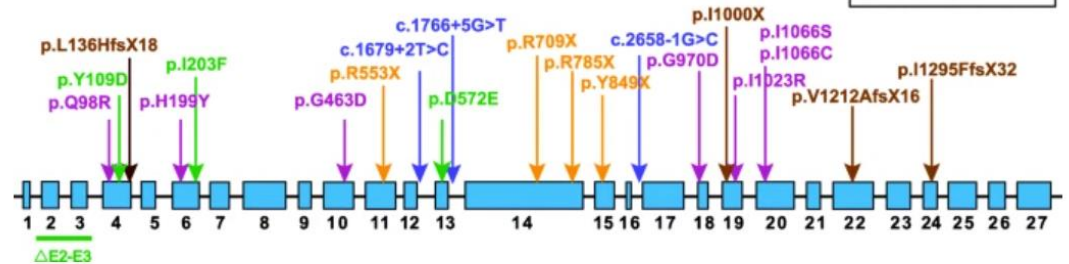
Purificazione e diluizione

- Utilizzo di biglie magnetiche per la purificazione del campione:
 - inserimento delle biglie all'interno del pool dell'amplificato
 - applicazione di un magnete
 - rimozione del surnatante
 - aggiunta di un buffer di lavaggio
 - aggiunta di un buffer di eluizione
 - applicazione del magnete
- Misurazione della concentrazione del DNA mediante fluorimetro.
- Diluizione del purificato (nM \rightarrow pM).



I primers

- i primers utilizzati per il sequenziamento di CFTR coprono tutte le regioni esoniche più circa 40 bp prima e dopo l'esone, per identificare anche mutazioni nelle regioni introniche.
 - sono circa 40
- nella reazione vengono anche inseriti dei primer specifici per il riconoscimento delle delezioni:
 - se c'è la delezione i due primer si trovano abbastanza vicini da poter amplificare una regione specifica.
 - se non c'è la delezione i due primer saranno "lontani" e quindi non in grado di amplificare.

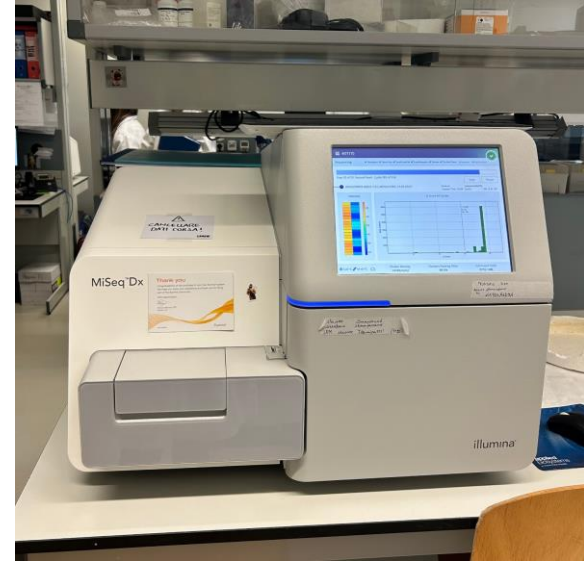


CFTR mutations detected in this CF cohort. Different mutation types are shown in the colors indicated in the upper panel; the gross deletion of exons 2–3 is indicated with a green solid line in the lower panel. The novel mutations identified in the current study are highlighted in green

NGS

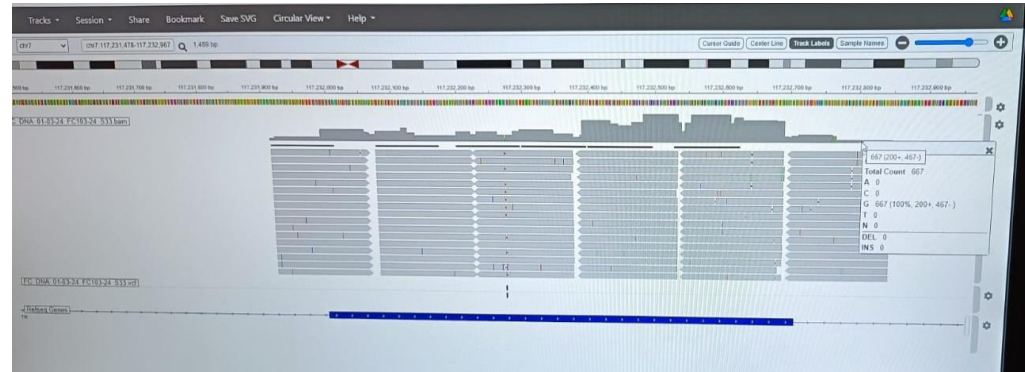
Q score= $-10\log(e)$

- Alto Q score-> minore probabilità di errore
- Basso Q score-> aumenta la probabilità di errore

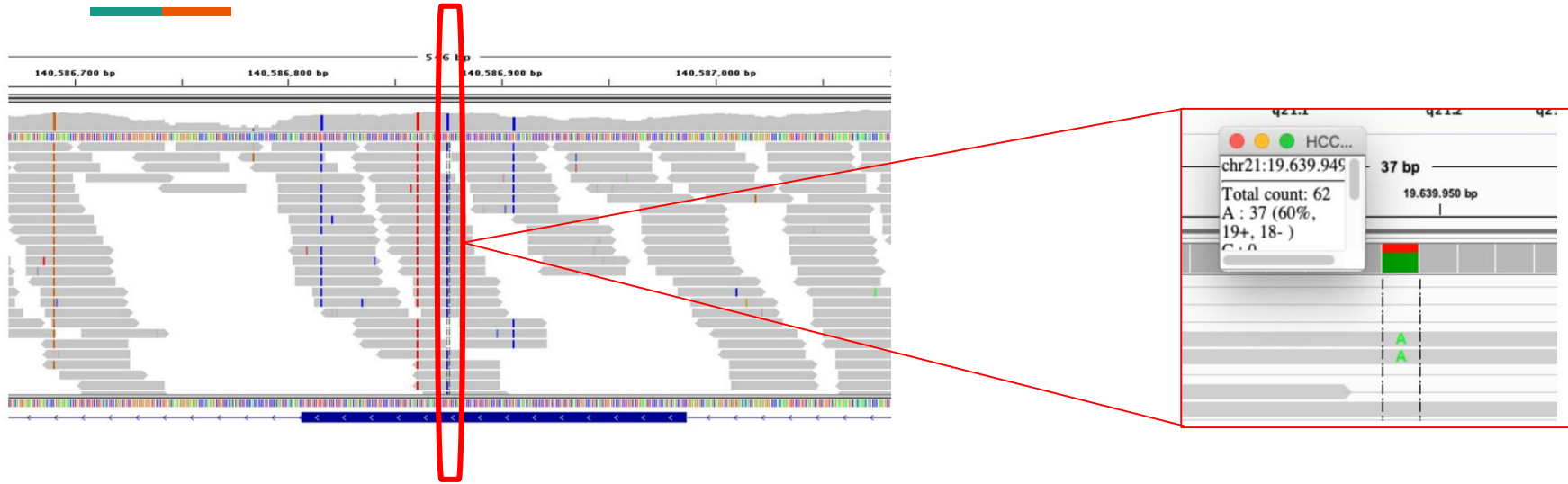


Le reads rappresentano le sequenze di ampliconi ottenute in PCR1 e PCR2.

- In grigio=sequenza della reads uguale al wild type
- Banda colorata= cambio di base



OMOZIGOSI VS ETEROZIGOSI



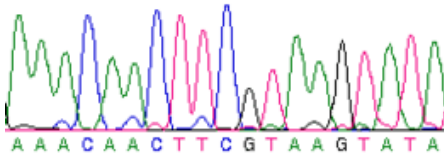
Su IGV si possono analizzare le sostituzioni e capire se la mutazione è in omo o in eterozigosi.

bisogna cliccare sulla sostituzione e il pop-up indicherà le percentuali che sono state riscontrate per ogni base: se una base è al 99% allora l'allele è in omozigosi, se invece abbiamo due basi circa attorno al 50% allora è in eterozigosi.

Sequenziamento Sanger

- PCR con terminazione di catena (tutti i ddNTP nella stessa miscela di reazione, ognuno legato a un marcatore fluorescente diverso)
- Corsa elettroforetica su un unico gel capillare
- Lettura da parte del computer delle bande del gel usando fluorescenza per identificare il ddNTP. Ognuno dei 4 ddNTP è dotato di un tag di fluorescenza diverso.

output: elettroferogramma.



se si ha un doppio picco c'è stato un cambio di base, se invece ho una delezione avrò un frameshift.



COME "DARE IL NOME" AD UNA MUTAZIONE: CFTR, NM_000492.4

HGVS

C. 2560 C>T

LEGACY

2428 C>T

C>T= sostituzione
Ala > Cis = sostituzione AA
C del = delezione
A ins = inserzione

C. = cerco la mutazione nel cDNA
G. = cerco la mutazione nel gene
p. = cerco la mutazione nella proteina

La prima base di trova a -132
bp dal codone d'inizio

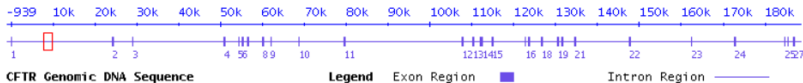
Posizione nucleotidica / AA all'interno del
gene / proteina dalla prima base A / dal
cordone d'inizio ATG

Cystic Fibrosis Mutation Database

[Home](#) [Search](#) [CFTR Gene](#) [History](#) [Team](#) [Statistics](#) [Links](#) [Submit](#) [Help](#)

← CFTR1

Select your region of interest by clicking on the graph below to obtain the corresponding CFTR genomic sequence spanning 2000 nucleotides



Enter the positions of the first and last nucleotides of the required CFTR sequence

From: To:

Move: Move Left Move right Don't Move

Zoom: Zoom In Zoom Out Don't Zoom

DNA sequence from 0 to 0 (Sequences corresponding to exons are shown in uppercase)

Get a sequence only copy. [DNA sequence](#)

Results for P5L and F508del

Variant P5L can be referred to as P5L, p.Pro5Leu, c.14C>T, or 146C>T

- The drug combination of tezacaftor and ivacaftor (Symdeko or Symkevi) and the drug combination of elexacaftor, tezacaftor, and ivacaftor (Trikafta or Kaftrio) have been approved in some countries for individuals with this variant. Please contact your physician to discuss whether these drugs are appropriate for you.

Variant F508del can be referred to as F508del, p.Phe508del, c.1521_1523delCTT, or c.1521_1523del or 1653delCTT,

- The drug combination of lumacaftor and ivacaftor (Orkambi), the drug combination of tezacaftor and ivacaftor (Symdeko or Symkevi), and the drug combination of elexacaftor, tezacaftor, and ivacaftor (Trikafta or Kaftrio) have been approved in some countries for individuals with this variant. Please contact your physician to discuss whether these drugs are appropriate for you.

CFTR2 →

<https://cftr2.org/mutation/general/P5L/F508del>

Summary Information

Sweat Chloride

Lung Function

% with Pancreatic Insufficiency

Pseudomonas Infection Rate

- This variant combination has varying consequences.
- Some patients with this variant combination have CF.
- Other patients with this variant combination do not have CF.
- Because of this variability, it is very important that CLINICAL CRITERIA ALONE be used to determine whether a person with this variant has CF.
- Because the clinical manifestations of CF can vary over the course of a person's lifetime, people who have this variant plus a variant that is known to cause CF should have periodic check-ups with their doctor even if they have no clinical signs or symptoms of CF at the present time.
- Clinical information shown below is taken only from patients in the CFTR2 database who have been diagnosed with CF.
 - There are other people with this variant who do NOT have CF. Information from these people is NOT included in the clinical information below, because these individuals are not followed at a CF center and are not part of the CFTR2 database. Therefore, the data below is not representative of every person with this variant.
- Patients with CF who have this variant are likely to be pancreatic sufficient.** This means they may not need to take oral pancreatic enzyme supplements every day.
- There are 27 patients with this variant combination in the CFTR2 database.

For help interpreting this information, we recommend you watch this video overview [What is Cystic Fibrosis?](#)

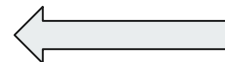
CFTR CF transmembrane conductance regulator [*Homo sapiens* (human)]

Download Datasets

Gene ID: 1080, updated on 23-Mar-2024

Summary

Official Symbol	CFTR provided by HGNC
Official Full Name	CF transmembrane conductance regulator provided by HGNC
Primary source	HGNC:HGNC:1884
See related	Ensembl:ENSG0000001626 MIM:602421 ; AllianceGenome:HGNC:1884
Gene type	protein coding
RefSeq status	REVIEWED
Organism	Homo sapiens
Lineage	Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi; Mammalia; Eutheria; Euarchontoglires; Primates; Haplorrhini; Catarrhini; Hominidae; Homo
Also known as	CF; MRP7; ABC35; ABCC7; CFTR/MRP; TNR-CFTR; dJ760C5.1
Summary	This gene encodes a member of the ATP-binding cassette (ABC) transporter superfamily. The encoded protein functions as a chloride channel, making it unique among members of this protein family, and controls ion and water secretion and absorption in epithelial tissues. Channel activation is mediated by cycles of regulatory domain phosphorylation, ATP-binding by the nucleotide-binding domains, and ATP hydrolysis. Mutations in this gene cause cystic fibrosis, the most common lethal genetic disorder in populations of Northern European descent. The most frequently occurring mutation in cystic fibrosis, DeltaF508, results in impaired folding and trafficking of the encoded protein. Multiple pseudogenes have been identified in the human genome. [provided by RefSeq, Aug 2017]
Expression	Biased expression in gall bladder (RPKM 28.8), colon (RPKM 22.3) and 6 other tissues See more
Orthologs	mouse all
NEW	Try the new Gene table
	Try the new Transcript table



PUBMED

[https://www.ncbi.nlm.nih.gov/clinvar/?term=CFTR\[gene\]](https://www.ncbi.nlm.nih.gov/clinvar/?term=CFTR[gene])

The Future of Genomic Medicine

Examples:

Enter variant, gene or select an example above

REFERENCE
hg19

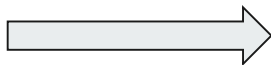
TYPE
Germline



FRANKLIN

<https://franklin.genoox.com/clinical-db/home>

MUTATION TASTER



<https://www.mutationtaster.org/>

Gene

Transcript

Position / snippet refers to
Alteration

Name of alteration

HGNC gene symbol, NCBI Gene ID, Ensembl gene ID [show available transcripts](#)

Ensembl transcript ID

Choose the transcript:

- [ENST0000003084](#) (*protein_coding*, 6128 bases) [NM_000492](#)
- [ENST00000454343](#) (*protein_coding*, 5949 bases)
- [ENST00000546407](#) (*processed_transcript*, 222 bases)
- [ENST00000472848](#) (*processed_transcript*, 148 bases)

coding sequence (ORF) transcript (cDNA sequence) gene (genomic sequence)

all types by sequence

enter a few bases around your alteration

Format:

ACTGTC[A/T] GTGTF A substituted by T
ACTGTC[AG/T] GTGTF AG substituted by T
ACTGTC[ACGT/] GTGTF ACGT deleted
ACTGTC[-AA] GTGTF AA inserted

single base exchange by position

enter position

and new base

insertion or deletion by position

enter positions of

...last wild type base before alteration

...first wild type base after alteration

and the inserted bases

(if applicable)

options

show nucleotide alignment

if you would like to have a name for this alteration in the output later on, please type in here

A 3D anatomical model of human lungs, showing the trachea and bronchial tree in a light orange color. The lungs are cut out of a light blue material. The model is surrounded by various paper-cut decorative elements, including a small orange butterfly, several light blue flowers, and a leaf. The background is a solid light blue color.

GRAZIE PER L'ATTENZIONE!